Resolution of Homoallylic Alcohols Containing Dithioketene Acetal Functionalities. Synthesis of Optically Active γ -Lactones by a Combination of Chemical and Enzymatic Methods

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Racemic homoallylic alcohols 1-3 containing dithioketeneacetal functionalities were prepared by addition of aldehydes to the allylic anions of ketene dithioacetals or 2-alkenyl-1,3-dithiane in a regio- and stereoselective manner. Lipase-catalyzed hydrolyses of the corresponding acetates 7–9 afforded optically active alcohols 1-3, which were treated with mercuric chloride to give γ -lactones such as natural hop lactone, whiskey lactone, and cognac lactone.

Introduction

Many natural products contain γ -lactone moieties. For example, γ -propenyl- γ -lactone is found in hop, $^{1}\beta$ -methyl- γ -butyl- γ -lactones (quercus lactones) are gradients of aged whiskey, $^{2}\beta$ -methyl- γ -pentyl- γ -lactones are natural flavor components of cognac,³ and β -methyl- γ -(3-methyl-2-butenyl)- γ -lactone (eldanolide) is a pheromone of the African sugarcane borer.⁴ We have synthesized such γ -lactones by hydrolyses of the homoallylic alcohols obtained by condensation of aldehydes with the allylic anions generated from ketene dithioacetals or 2-propenyl-1,3-dithiane (4-6).⁵ Advantageously, the reaction is γ -regioselective and the relative stereochemistry of the β - and γ -substituents in **3** is controlled. Racemic **1**-**3** can be kinetically resolved using lipase-catalyzed reactions, and the optically active homoallylic alcohols containing dithioketene acetal functionalities can be converted to γ -lactones such as natural hop lactone, whiskey lactone, and cognac lactone.

Results and Discussion

The allylic anion generated from the ketene dithioacetal (4 or 5) or the 2-propenyl-1,3-dithiane (6) in THF was treated with a variety of aldehydes to give the homoallylic alcohols 1-3. Compounds 3 had the three configuration as they were obtained via chairlike sixmembered cyclic transition states.⁵ A number of lipases, such as lipases AP6, MY, AY-30, OF, AK, PS, M-AP10, CE-10, R-10, GC-4, N, and type 1 as well as porcine pancreas lipase were tested in order to resolve the homoallylic alcohols 1. Acetylation of some homoallylic alcohols (Table 1) can be conducted in the presence of vinyl acetate by catalysis with lipases MY or AY, though

* Abstract published in Advance ACS Abstracts, September 1, 1994. (1) Naya, Y.; Kotake, M. Inst. Food. Chem., Osaka, Japan 1968, 89, 1113.

R ^{AC} OH	R - OAc
1a-s	7a-s
1a R = Me 1b R = Et 1c R = r -Pr 1d R = λ Pr 1e R = C ₆ H ₅ 1f R = σ -MeC ₆ H ₄ 1g R = m -MeC ₆ H ₄ 1h R = p -MeC ₆ H ₄ 1i R = p -(λ Pr)C ₆ H ₄ 1j R = σ -MeOC ₆ H ₄ 1k R = m -MeOC ₆ H ₄ 1k R = m -CF ₃ C ₆ H ₄ 1k R = m -CF ₃ C ₆ H ₄ 1m R = p -CF ₃ C ₆ H ₄ 1m R = p -CF ₃ C ₆ H ₄ 1m R = m -CF ₃ C ₆ H ₄ 1m R = m -CC ₆ H ₄ 1m R = m -CC ₆ H ₄ 1m R = m -BrC ₆ H ₄ 1m	7a R = Me 7b R = Et 7c R = <i>n</i> -Pr 7d R = <i>i</i> -Pr 7e R = C_6H_5 71 R = <i>o</i> -MeC_6H_4 7g R = <i>m</i> -MeC_6H_4 7h R = <i>p</i> -(<i>i</i> -Pr)C_6H_4 7j R = <i>o</i> -MeOC_6H_4 7k R = <i>m</i> -MeOC_6H_4 7k R = <i>m</i> -CF_3C_6H_4 7n R = <i>p</i> -CF_3C_6H_4 7n R = <i>p</i> -FC_6H_4 7p R = <i>o</i> -CIC_6H_4 7p R = <i>m</i> -CIC_6H_4
R OH	R OAc
2a-g	8a-g
2a R = Me 2b R = Et 2c R = r -Pr 2d R = i -Pr 2e R = C_6 H ₅ 2f R = r -Bu 2g R = r - C_5 H ₁₁	8a R = Me 8b R = Et 8c R = n -Pr 8d R = i -Pr 8e R = C_6H_5 8f R = n -Bu 8g R = n - C_5H_{11}
Me R ^{····} OH 3a-d	R OAc 9a-d
3a R = Et 3b R = r -Bu 3c R = r -C ₅ H ₁₁ 3d R = r -C ₆ H ₁₃	9a R = Et 9b R = <i>n</i> -Bu 9c R = <i>n</i> -C ₅ H ₁₁ 9d R = <i>n</i> -C ₆ H ₁₃

the enantioselectivity was usually low. Acetylations of 1d, 2d, and 2e by catalysis with lipase MY (entries 6,

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[†] Institute of Biological Chemistry.

<sup>(1113.)
(2) (</sup>a) Masuda, M.; Nishimura, K. Phytochemistry 1971, 10, 1401.
(b) Masuda, M.; Nishimura, K. Chem. Lett. 1981, 1333. (c) Hoppe, D.;
Bronneke, A. Tetrahedron Lett. 1983, 24, 1687.
(3) (a) Ortuno, R. M.; Merce, R.; Font, J. Tetrahedron 1987, 43, 4497.

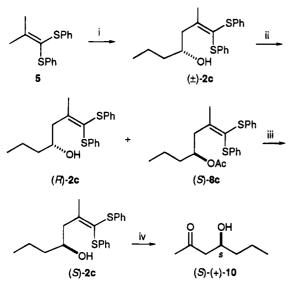
⁽b) Nishimura, K. Chem. Today 1987, 189, 30. (c) Ebata, T.; Mastsu-moto, K; Yoshikoshi, H.; Koseki, K.; Kawakami, H.; Matsushita, H.

^{moto, K; Yoshikoshi, H.; Koseki, K.; Kawakami, H.; Matsushita, H.} Hetercoycles 1990, 31, 1585.
(4) Kunesch, G.; Zagatti, P.; Lallemand, J. Y.; Debal, A.; Vigneron, J. P. Tetrahedron Lett. 1981, 22, 5271.
(5) (a) Fang, J.-M.; Hong, B.-C. Synth. Commun. 1986, 16, 523. (b) Fang, J.-M.; Liao, L.-F.; Hong, B.-C. J. Org. Chem. 1986, 51, 2828. (c) Fang, J.-M.; Hong, B.-C.; Liao, L.-F. J. Org. Chem. 1987, 52, 855

Table 1. Acetylation of Homoallylic Alcohols 1 and 2 with Vinyl Acetate in Hexane by Catalysis with Lipases

entry	alcohol substrate	R =	lipase	reaction time (h)	conversn (%)	acetate product	ee _p (%)	confign of major enantiomer	ee _s (%) of remaining substrate	E
1	1a	Me	MY	3	80	7a	8	S	8	1.3
2	1 a	Me	AY	1	50	7a	6	\boldsymbol{s}	6	1.2
3	1b	Et	MY	13	50	7b	4	\boldsymbol{s}	4	1.1
4	1b	Et	AY	9	50	7b	12	\boldsymbol{S}	26	1.5
5	1c	n-Pr	MY	25	50	7c	6	\boldsymbol{S}	16	1.3
6	1 d	<i>i</i> -Pr	MY	150	20	7d	93	R	16	32
7	2a	Me	MY	12	50	8a	40	\boldsymbol{s}	40	3.3
8	2a	Me	AY	12	50	8a	40	\boldsymbol{S}	39	3.4
9	2b	\mathbf{Et}	MY	48	50	8b	46	\boldsymbol{S}	52	4.4
10	2b	\mathbf{Et}	AY	48	47	8b	69	\boldsymbol{S}	69	11
11	2c	n-Pr	MY	100	25	8c	73	\boldsymbol{S}	24	8
12	2c	n-Pr	AY	25	44	8c	86	\boldsymbol{S}	67	27
13	2d	<i>i</i> -Pr	MY	150	13	8 d	96	R	22	60
14	2e	C_6H_5	MY	240	10	8e	98	R	15	114

Scheme 1^a



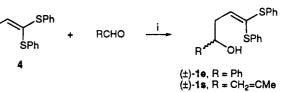
^a Key: (i) BuLi, THF, n-C₃H₇CHO, -78 °C, 10 min; 85%; (ii) lipase, vinyl acetate (10 equiv), hexane, see Table 1; (iii) aqueous KOH (30%), MeOH, 25 °C, 3 h; 98%; (iv) O₃, CH₂Cl₂, -78 °C; Me₂S, 25 °C.

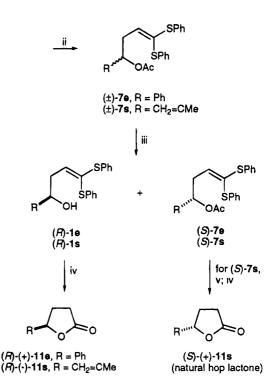
13, and 14, R = i-Pr or Ph) were sluggish but gave acetates 7d, 8d, and 8e in high optical purity ($\geq 93\%$ ee).

Reaction mixture components were separated by silica gel chromatography, and the enantiomeric excess of the remaining substrate, ees, and the product, eep, were determined by HPLC using a Chiracel OD column. The E value was calculated according to $E = \ln[1 - c(1 + c)]$ ee_p]/ln[1-c(1 - ee_p)],⁶ where conversion, c, was deduced from analysis of the reaction mixture by GC, HPLC, or ¹H NMR spectroscopy. The acetate 8c (86% ee), obtained by the lipase-catalyzed reaction (entry 12, Table 1), was saponified and ozonized to give (+)-4-hydroxy-2-heptanone⁷ in favor of the (S)-configuration (Scheme 1).

Alternatively, lipase-catalyzed hydrolyses of racemic acetates 7 and 8 were carried out in the presence of cosolvents (Tables 2 and 3). The rate of hydrolysis of 7e (R = Ph) by lipase MY was adequate using either DMF, hexane, or toluene. The hydrolysis in phosphate buffer (pH 7.5) with 10% DMF appeared to be superior, giving alcohol 1e in higher enantioselectivity (compare entries 3-5, Table 2).

Scheme 2^a





^a Key: (i) BuLi, THF, -78 °C, 10 min; 85%; (ii) Ac₂O, Et₃N, DMAP cat., CH₂Cl₂, 25 °C, 1 h; 98%; (iii) lipase, DMF, phosphate buffer (pH 7.5); see Table 2; (iv) HgCl₂, aqueous MeOH (10%), reflux, 5h; 85%; (v) aqueous KOH (30%), MeOH, 25 °C, 3 h; 98%.

Compounds **7f-r** containing either electron-donating or electron-withdrawing groups on the phenyl ring were substrates for the lipase-catalyzed reaction. Lipases MY and AY catalyzed the hydrolyses of acetates 7d-s in the same stereospecific manner. The major enantiomer of alcohol products 1d-s in each case has the longer retention time on the Chiracel OD column. As delineated in Scheme 2, lipase-catalyzed hydrolyses gave homoallylic alcohols 1e (R = Ph) and $1s (R = CH_2=CMe)$ in favor of the (R)-configuration. Product 1e (91% ee) was treated with $HgCl_2$ to give the optically active γ -phenyl- γ -lactone **11e**,⁸ $[\alpha]_D$ +18° (c = 1.1, CHCl₃). The (R)-enantiomer of

⁽⁶⁾ Chen, C.-J.; Fujimoto, Y.; Girdaukas, G.; Sih, C. J. J. Am. Chem. Soc. 1982, 104, 7294

⁽⁷⁾ Narasaka, K.; Miwa, T.; Hayashi, H.; Ohta, M. Chem. Lett. 1984, 1399.

Table 2. Lipase-Catalyzed Hydrolysis of Acetates 7 in Organic Solvent/Phosphate Buffer (pH 7.5) Solution

entry	acetate substrate	R =	lipase	organic solvent	reaction time (h)	conversn (%)	alcohol product	ee _p (%)	confign of major enantiomer	ee _s (%) of remaining substrate	E
1	7d	<i>i</i> -Pr	MY	hexane	240	35	1d	95	R	50	58
2	7d	<i>i</i> -Pr	MY	DMF	216	42	1d	95	R	68	72
3	7e	C_6H_5	MY	hexane	120	37	1e	79	R	45	13
4	7e	C_6H_5	MY	toluene	300	50	1e	75	R	99	34
5	7e	C_6H_5	MY	\mathbf{DMF}	120	50	1e	82	R	87	33
6	7e	C_6H_5	AY	DMF	168	41	1e	91	R	64	40
7	7f	$o-MeC_6H_4$	MY	DMF	120	20	1f	93	R	27	35
8	7g	m-MeC ₆ H ₄	MY	DMF	130	30	1g	91	R	45	33
9	7g	m-MeC ₆ H ₄	AY	\mathbf{DMF}	210	32	1g	87	R	47	23
10	7h	$p-MeC_6H_4$	MY	\mathbf{DMF}	160	44	1 h	87	R	71	30
11	7i	$p-(i-\Pr)C_6H_4$	MY	\mathbf{DMF}	288	45	1i	94	R	70	64
12	7j	$o-MeOC_6H_4$	MY	\mathbf{DMF}	120	35	1j	92	R	50	39
13	7k	m-MeOC ₆ H ₄	MY	DMF	120	23	1k	96	R	37	70
14	71	m-CF ₃ C ₆ H ₄	MY	DMF	130	40	11	98	R	51	164
15	71	m-CF ₃ C ₆ H ₄	AY	DMF	120	20	11	97	R	19	79
16	7m	p-CF ₃ C ₆ H ₄	MY	DMF	144	28	1m	96	R	28	70
17	7n	$m-\mathrm{FC}_6\mathrm{H}_4$	MY	\mathbf{DMF}	120	38	1 n	94	R	56	57
18	70	$p-\mathrm{FC}_6\mathrm{H}_4$	MY	DMF	120	40	1o	96	R	54	113
19	7p	$o-ClC_6H_4$	MY	\mathbf{DMF}	130	33	1p	98	R	47	125
20	$7\overline{q}$	$m-ClC_6H_4$	MY	DMF	120	48	1q	97	R	84	190
21	$7\bar{q}$	m-ClC ₆ H ₄	AY	DMF	120	42	1q	89	R	66	34
22	7r	m-BrC ₆ H ₄	MY	DMF	310	20	1r	81	R	20	11
23	7s	$CH_2 = CMe$	MY	DMF	120	37	1s	82	R	42	13
24	7s	$CH_2 = CMe$	AY	DMF	240	55	1s	75	R	95	222

Table 3. Lipase-Catalyzed Hydrolysis of Acetates 8 in DMF/Phosphate Buffer (pH 7.5) Solution (1:9)

entry	acetate substrate	R =	lipase	reaction time (h)	conversn (%)	alcohol product	ee _p (%)	confign of major enantiomer	ee _s (%) of remaining substrate	E
1	8a	Me	MY	24	50	2a	23	S	21	2
2	8a	Me	AY	24	70	2a	20	\boldsymbol{S}	37	4.4
3	8b	Et	MY	120	43	2b	56	\boldsymbol{S}	31	4
4	8b	\mathbf{Et}	AY	120	48	2b	57	\boldsymbol{S}	54	6
5	8c	n-Pr	MY	120	45	2c	61	\boldsymbol{S}	40	6
6	8c	<i>n</i> -Pr	AY	120	40	2c	65	\boldsymbol{S}	36	7
7	8 d	<i>i</i> -Pr	MY	450	30	2d	77	R	36	11
8	8d	<i>i</i> -Pr	AY	360	35	2d	91	R	55	37
9	8e	C_6H_5	MY	144	47	2e	66	R	53	8
10	8e	C_6H_5	AY	48	25	2e	74	R	24	8
11	8f	n-Bu	MY	137	45	2f	72	\boldsymbol{S}	48	10
12	8f	n-Bu	AY	137	45	2f	82	\boldsymbol{S}	67	20
13	8g	$n-C_5H_{11}$	MY	220	15	2g	97	\boldsymbol{S}	21	120
14	8g	n-C ₅ H ₁₁	AY	216	15	2g	99	\boldsymbol{S}	12	220

Table 4. Lipase-Catalyzed Hydrolysis of Acetates 9 in DMF/Phosphate Buffer (pH 7.5) Solution (1:9)

entry	acetate substrate	R =	lipase	reaction time (h)	conversn (%)	alcohol product	ee p (%)	confign of major enantiomer	ee _s (%) of remaining substrate	E
1	9a	Et	MY	48	40	3a	88	2S,3R	55	27
2	9a	\mathbf{Et}	AY	48	38	3a	91	2S, 3R	52	35
3	9b	n-Bu	MY	70	30	3b	91	2S, 3R	44	33
4	9b	<i>n-</i> Bu	AY	70	25	3b	98	2S, 3R	35	139
5	9b	n-Bu	AY	480	53	3b	85	2S, 3R	95	48
6	9c	n-C ₅ H ₁₁	MY	144	40	3c	85	2S, 3R	58	22
7	9c	$n \cdot C_5 H_{11}$	AY	51	37	3c	93	2S, 3R	65	52
8	9c	$n-C_5H_{11}$	AY	240	24	3c	96	2S, 3R	30	66
10	9d	$n-C_{6}H_{13}$	MY	216	40	3d	81	2S, 3R	58	17
10	9d	$n-C_6H_{13}$	AY	216	30	3d	94	2S, 3R	37	46

alcohol **1s** (95% ee) was similarly converted to (-)-hop lactone **11s**. The natural hop lactone is dextrorotatory¹ and by these data has the (S)-configuration.

The combined chemical and enzymatic method was used in an expedient synthesis of a whisky lactone **12b** and a cognac lactone **12c** containing trans β - and γ -substituents (Scheme 3). The racemic alcohol **3b** (R = n-C₄H₉) obtained by condensation of 2-propenyl-1,3dithiane and pentanal was converted to the correspond-

(8) Manzocchi, A.; Rosangela, C.; Fiecchi, A.; Santaniello, E. J. Chem. Soc., Perkin Trans. 1 1987, 2753.

ing acetate **9b** (Ac₂O, Et₃N). Racemic **9b** was subjected to lipase-catalyzed hydrolysis to give the optically active alcohol **3b** having the (2S,3R)-configuration (Table 4). Subsequent treatment with HgCl₂ afforded the natural trans quercus lactone **12b**.² By a similar procedure, the optically active alcohol **3c** (R = n-C₅H₁₁) was obtained and subsequently transformed into the natural cognac lactone **12c**.³ The remaining acetates (2*R*,3*S*)-**9b**,**c** were saponified and treated with HgCl₂ to give unnatural antipodes of whiskey lactone and cognac lactone.

In summary, we have demonstrated the synthesis of varied optically active alcohols 1-3 via lipase-catalyzed

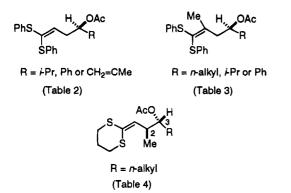
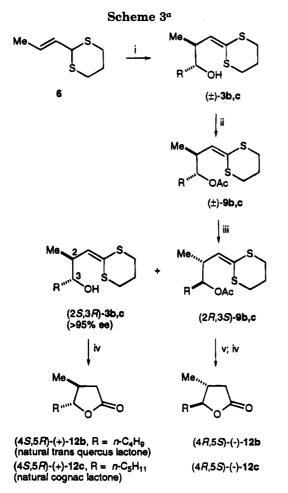


Figure 1. Preferable enantiomers for lipase-catalyzed hydrolyses.



^a Key: (i) BuLi, THF, n-C₄H₉CHO or n-C₅H₁₁CHO, -78 °C, 10 min; 85%; (ii) Ac₂O, Et₃N, cat. DMAP, CH₂Cl₂, 25 °C, 1 h; 98%; (iii) lipase, DMF, phosphate buffer (pH 7.5), see Table 4; (iv) HgCl₂, aqueous MeOH (10%), reflux, 5 h; 92%.

hydrolyses of racemic acetates 7–9. Alcohols 1s, 3b, and 3c are precursors of natural γ -lactones. The enantiomeric preference for lipase-catalyzed hydrolyses of 7–9 are shown in Figure 1. (*R*)-Enantiomers of 7d (R = *i*-Pr), 7e-r (R = XC₆H₄), and 7s (R = CH₂=CMe) were selectively hydrolyzed with 75–98% ee (Table 2). The result indicates a model for the enzymatic hydrolysis. The molecule orients with the bis(phenylthio) group on the left-hand side and the R group on the right-hand side, while the reacting acetoxy group positions on the front face. A similar orientation of substrates 8, regardless of the nature of R groups (*n*-alkyl, *i*-Pr, or Ph) also accounts for the observed enantioselective hydrolyses (Table 3). Lipase-catalyzed acetylations of alcohols 1 and 2 (Table 1) follow the same trend, i.e., preference for formation of (R)-enantiomers of acetates **7a**, **8d**, and **8e** ($\mathbf{R} = i$ -Pr or Ph) and (S)-enantiomers of **7a-c** and **8a-c** ($\mathbf{R} = n$ -alkyl). Lipase-catalyzed hydrolysis of **9**, however, appears to favor the (2S,3R)-enantiomer having the molecule oriented as shown in Figure 1. From the literature,⁹ alcohols containing sulfanyl or sulfonyl groups have been resolved by enzymatic methods. Several models¹⁰ have been proposed to interpret enantioselectivity in lipase-catalyzed reactions, though no single model can fit all the experimental results.

Experimental Section

The ¹H NMR spectra were recorded at 200 or 300 MHz using tetramethylsilane as internal standard. ¹³C NMR spectra were recorded at 50 or 75 MHz. The mass spectra were recorded at an ionizing voltage of 70 or 20 eV. HPLC was carried out on a chromatograph using a μ -Porasil column (7 μ m, 25 cm \times 0.78 cm) with a 5 mL/min flow rate of elution. Enantiomeric excess of the remaining substrate, ee_s, and the product, ee_p, were determined by HPLC using a Chiracel OD column (0.46 cm i.d. \times 25 cm) with 1 mL/min flow rate of elution. The *E* value was calculated according to $E = \ln[1 - c(1 + ee_p)]/\ln[1 - c(1 - ee_p)],^6$ where *c* is conversion.

Lipase AP6 (Aspergillus niger), lipase PS (Pseudomonas sp.), lipase N (Rhizopus niveus), lipase F-AP15 (Rhizopus oryzae), lipase AY-30 (Candida cylindracea), lipase CE-10 (Humicola sp.), lipase GC-4 (Geotrichum candidum), lipase R-10 (Penicillium roqueforti), lipase AK (Pseudomonas sp.), and lipase M-AP10 (Mucor meihei), were purchased from Amano Pharm. ipase OF (C. cylindracea) and lipase MY (C. cylindracea) were from Meito-Sangyo Co., Ltd, Japan. Porcine pancreas lipase (PPL) and lipase type 1 (wheat) were from Sigma, USA. These crude enzymes were used for enzymatic reactions without further purification.

General Procedure for Synthesis of Racemic Homoallylic Alcohols 1-3 and Their Acetates 7-9. Under an atmosphere of nitrogen, butyllithium (4.3 mL of 1.6 M solution in hexane) was added drop by drop to a cold (-20 °C) THF (10 mL) solution of dithioketene acetal or dithiane (5 mmol, 4, 5, or 6). The mixture was stirred for 30 min and cooled to -78 °C, and a THF (2 mL) solution of the appropriate aldehyde (6 mmol) was added drop by drop. The mixture was stirred for 10 min and quenched by addition of a solution of acetic acid (0.9 mL, 15 mmol) in THF (2 mL). The mixture was washed with saturated NaHCO3 and extracted three times with EtOAc. The combined EtOAc extracts were washed with brine, dried (Na_2SO_4) , and concentrated under reduced pressure. The residue was chromatographed on a silica gel column by elution with gradients of EtOAc in hexane (10-20%) to give homoallylic alcohols 1-3 in 85-95% yields.

Triethylamine (10 mmol) and acetic anhydride (16 mmol) were added to a cold (0 $^{\circ}$ C) solution of the homoallylic alcohol

(10) (a) Oberhauser, T.; Faber, K.; Griengl, H. Tetrahedron 1989, 45, 1679. (b) Roberts, S. M. Phill. Trans. R. Soc. London B 1989, 45, 1679. (c) Xie, Z.-F.; Suemune, H.; Sakai, K. Tetrahedron Asymmetry 1990, 1, 395. (d) Burgess, K.; Jennings, L. D. J. Am. Chem. Soc. 1990, 112, 7434. (e) Kazlauskai, R. J.; Weissfoch, A. N. E.; Rappaport, A. T.; Cuccia, L. A. J. Org. Chem. 1991, 56, 2656 and references cited therein. (f) Burgess, K.; Jennings, L. D. J. Am. Chem. Soc. 1991, 113, 6129. (g) Kim, M.-J.; Cho, H. J. Chem. Soc., Chem. Commun. 1992, 1411. (h) O'Hagan, D.; Zaid, N. A. J. Chem. Soc., Perkin Trans. I 1992, 947. (i) Wimmer, Z. Tetrahedron 1992, 48, 8431. (j) Takano, S.; Yamane, T.; Takahashi, M.; Ogasawara, K. Tetrahedron Asymmetry 1992, 3, 837.

^{(9) (}a) Fuganti, C.; Grasselli, P.; Servi, S.; Lazzarine, A.; Casati, P. Tetrahedron 1988, 44, 2575. (b) Bianchi, D.; Cesti, P.; Golini, P. Tetrahedron 1989, 45, 869. (c) Gu, R. L.; Sih, C. J. Tetrahedron Lett. 1990, 31, 3283. (d) Jacobs, H. K.; Mueller, B. H.; Gopalan, A. S. Tetrahedron 1992, 48, 8891. (e) Goergens, U.; Schneider, M. P. Tetrahedron Asymmetry 1992, 3, 1149. (f) Oyama, K.; Nishimura, S.; Nonaka, Y.; Kihara, K.; Hashimoto, T. J. Org. Chem. 1981, 46, 5242. (g) Isowa, Y.; Ohmori, M.; Ichikawa, T.; Mori, K. Tetrahedron Lett. 1979, 2611. (h) Sakina, K.; Kawazura, K.; Morihara, K.; Tajima, H. Chem. Pharm. Bull. 1988, 36, 4345. (i) Matos, J. R.; West, J. B.; Wong, C.-H. Biotechnol. Lett. 1977, 9, 233.

(4 mmol) and a small amount of 4-(dimethylamino)pyridine (10 mg) in CH₂Cl₂ (10 mL). The mixture was stirred for 1 h, concentrated, diluted with brine, and extracted with EtOAc. The organic phase was dried (Na₂SO₄), concentrated, and purified on a silica gel column by elution with gradients of EtOAc in hexane (3-10%) to give the corresponding acetates **7-9** in 90-98% yields.

General Procedure for Lipase-Catalyzed Acetylation of Homoallylic Alcohols 1 and 2. The alcohol (1 mmol) was stirred (800-1000 rpm) with vinyl acetate (10 mmol) and a lipase (0.3 g) in hexane (5 mL) at room temperature (25-27 °C). An aliquot of the reaction mixture was occasionally taken and filtered, and the filtrate was analyzed by GC, HPLC, or ¹H NMR to determine the conversion. After the mixture was stirred for the period indicated in Table 1, 30-55% of the alcohol was converted to the corresponding acetate. The mixture was filtered, and the filtrate was concentrated and chromatographed on a silica gel column by elution with gradients of EtOAc in hexane (3-15%) to give optically active acetate and alcohol. Enantiomeric excess of the remaining alcohol was determined by HPLC using a Chiracel OD column with elution of 2-propanol in hexane (0.5-10%). In order to determine the optical purity of the product, the acetate was converted to the corresponding alcohol by saponification in aqueous KOH (30%, 1 mL)/MeOH (10 mL) at room temperature for 3 h. The optical purity of the alcohol was similarly determined by HPLC, and its value was taken as that of the acetate.

General Procedure for Lipase-Catalyzed Hydrolysis of Racemic Acetates 7–9. The acetate (1 mmol) and a lipase (0.3 g) in a mixed solvent of DMF (0.8 mL) and phosphate buffer (7.2 mL, pH 7.5) were stirred (800–1000 rpm) at room temperature (25–27 °C) for the period indicated in Tables 2–4 to reach 30–55% conversion. The mixture was filtered, and the filtrate was analyzed by GC, HPLC, or ¹H NMR to determine the percent conversion to product. The alcohol products 1–3 and the starting materials 7–9 were separated by chromatography, and their values of optical purity were determined by HPLC as described above. In the entries 1, 3, and 4 of Table 2, the ratio of cosolvent hexane (or toluene) to phosphate buffer was 1:3.

The physical and spectral data of 3-6 were previously reported.^{5,11}

5,5-Bis(phenylthio)-4-penten-2-ol (1a): oil; TC (15% EtOAc in hexane) $R_f = 0.25$; IR (neat) 3355 cm⁻¹; MS m/z (rel intensity) 302 (M⁺, 100), 257 (90); ¹H NMR (CDCl₃) δ 7.28–7.18 (10 H, m), 6.37 (1 H, t, J = 7.4 Hz), 3.90 (1 H, dq, J = 6.2, 6.0 Hz), 2.60 (2 H, dd, J = 7.2, 7.8 Hz), 1.21 (3 H, d, J = 6.2 Hz); ¹³C NMR (CDCl₃) δ 138.6 (d), 136.0 (s), 135.0 (s), 133.9 (s), 131.6 (d, 2 C), 130.4 (d, 2 C), 128.8 (d, 2 C), 128.7 (d, 2 C), 127.4 (d), 126.8 (d), 67.4 (d), 40.6 (t), 23.2 (q); HRMS calcd for C₁₇H₁₈OS₂ (M⁺) 302.0799, found 302.0801; HPLC (Chiracel OD, 2-propanol/hexane (3:97)) $t_{\rm R}$ 15.8 min (*R*-isomer), 18.7 min (*S*-isomer).

6,6-Bis(phenylthio)-5-hexen-3-ol (1b): oil; ¹³C NMR (CDCl₃) δ 139.3 (d), 134.0 (s, 2 C), 131.6 (d, 2 C), 131.5 (s), 130.4 (d, 2 C), 128.8 (d, 2 C), 128.7 (d, 2 C), 127.4 (d), 126.8 (d), 72.7 (d), 38.6 (t), 30.0 (t), 9.5 (q); HRMS calcd for C₁₈H₂₀-OS₂ (M⁺) 316.0955, found 316.0955; HPLC (Chiracel OD, 2-propanol/hexane (3:97)) $t_{\rm R}$ 18.7 min (*R*-isomer), 21.9 min (*S*-isomer).

1,1-Bis(phenylthio)-1-hepten-4-ol (1c): oil; ¹³C NMR (CDCl₃) δ 139.3 (d), 134.0 (s, 2 C), 131.6 (d, 4 C), 130.4 (d), 128.8 (d, 2 C), 128.7 (d), 127.4 (d), 126.8 (d), 71.0 (d), 39.3 (t), 39.1 (t), 18.7 (q); HRMS calcd for C₁₉H₂₂OS₂ (M⁺) 330.1112 Found 330.1118; HPLC (Chiracel OD, 2-propanol/hexane (5: 95)) t_R 11.9 min (*R*-isomer), 14.5 min (*S*-isomer).

6,6-Bis(phenylthio)-2-methyl-5-hexen-3-ol (1d): oil; ¹³C NMR (CDCl₃) δ 140.1 (d), 134.0 (s), 131.4 (s), 130.9 (s), 130.4 (d, 2C), 128.7 (d, 2 C), 128.6 (d, 2 C), 127.3 (d), 126.7 (d), 76.1 (d), 36.2 (t), 33.5 (d), 18.7 (q), 17.2 (q); HRMS calcd for C₁₉H₂₂-

 OS_2 (M⁺) 330.1112, found 330.1112; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) t_R 8.1 min (S-isomer), 10.7 min (R-isomer).

4,4-Bis(phenylthio)-1-phenyl-3-butenol (1e): oil; ¹³C NMR (CDCl₃) δ 143.4 (s), 138.0 (d), 134.0 (s), 133.8 (s), 131.7 (d, 2 C), 130.4 (d, 2 C), 128.8 (d, 4 C), 128.6 (s), 128.5 (d, 2 C), 127.6 (d), 127.4 (d), 126.8 (d), 125.8 (d, 2 C), 75.6 (d), 40.6 (t); HRMS calcd for C₂₂H₂₀OS₂ (M⁺) 364.0956, found 364.0961; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 15.9 min (S-isomer), 17.4 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(o-methylphenyl)-3-butenol (1f): oil; ¹³C NMR (CDCl₃) δ 141.5 (s), 138.2 (d), 134.4 (s), 134.1 (s), 134.0 (s), 133.7 (s), 131.8 (d, 2 C), 130.4 (d), 130.3 (d, 2 C), 128.8 (d, 2 C), 128.6 (d, 2 C), 127.4 (d), 127.3 (d), 126.7 (d), 126.3 (d), 125.3 (d), 70.0 (d), 39.3 (t), 19.0 (q); HRMS calcd for C₂₃H₂₂OS₂ (M⁺) 378.1112, found 378.1086; HPLC (Chiracel OD, 2-propanol/hexane (20:80)) $t_{\rm R}$ 5.9 min (S-isomer), 9.6 min (R-isomer).

4,4-Bis(phenylthio)-1-(m-methylphenyl)-3-butenol (1g): oil; ¹³C NMR (CDCl₃) δ 143.4 (s), 138.4 (s), 138.0 (s), 134.0 (s), 133.8 (s), 131.6 (d, 2 C), 130.3 (d, 2 C), 128.7 (d, 2 C), 128.6 (d, 2 C), 128.3 (d, 2 C), 127.3 (d, 2 C), 126.7 (d), 126.4 (d), 122.9 (d), 73.5 (d), 40.5 (t), 21.4 (q), 14.1 (q); HRMS calcd for C₂₃H₂₂-OS₂ (M⁺) 378.1112, found 378.1112; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 9.3 min (S-isomer), 10.5 min (Risomer).

4,4-Bis(phenylthio)-1-(p-methylphenyl)-3-butenol (1h): oil; ¹³C NMR (CDCl₃) δ 140.5 (s), 138.6 (d), 137.2 (d), 135.9 (s), 135.6 (s), 134.0 (s), 133.8 (s), 131.6 (d, 2 C), 131.5 (d, 2 C), 128.9 (d), 128.7 (d, 2 C), 128.5 (d, 2 C), 127.3 (d), 126.7 (d), 125.8 (d, 2 C), 73.4 (d), 40.1 (t), 21.0 (q); HRMS calcd for C₂₃H₂₂-OS₂ (M⁺) 378.1112, found 378.1112; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 14.8 min (S-isomer), 17.6 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(p-(methylethyl)phenyl)-3-butenol (1i): oil; ¹³C NMR (CDCl₃) δ 148.4 (s), 140.9 (d), 138.3 (d), 134.0 (d), 133.8 (d), 131.7 (d, 2 C), 131.7 (s), 130.4 (d, 2 C), 128.7 (d, 2 C), 128.6 (d, 2 C), 127.4 (d), 126.7 (d), 126.5 (d, 2 C), 125.8 (d, 2 C), 73.5 (d), 40.5 (t), 33.8 (d), 24.0 (q, 2 C); HRMS calcd for C₂₅H₂₆OS₂ (M⁺) 406.1425, found 406.1419; HPLC (Chiracel OD, 2-propanol/hexane (20:80)) $t_{\rm R}$ 10.4 min (S-isomer), 14.0 min (R-isomer).

4,4-Bis(phenylthio)-1-(o-methoxyphenyl)-3-butenol (1j): oil; ¹³C NMR (CDCl₃) δ 156.5 (s), 139.8 (d), 134.3 (s), 134.1 (s), 131.5 (d, 2 C), 131.2 (s), 131.0 (s), 130.3 (d, 2 C), 128.7 (d, 2 C), 128.6 (d, 2 C), 128.5 (d), 127.2 (d),127.1 (d), 126.6 (d), 120.8 (d), 110.6 (d), 70.6 (d), 55.3 (q), 39.0 (t); HRMS calcd for C₂₃H₂₂O₂S₂ (M⁺) 394.1061, found 394.1067; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 14.4 min (S-isomer), 18.4 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(m-methoxyphenyl)-3-butenol (**1k**): oil; ¹³C NMR (CDCl₃) δ 159.8 (s), 145.2 (s), 138.0 (d), 134.0 (s), 133.8 (s), 132.0 (s), 131.8 (d, 2 C), 130.4 (d, 2 C), 129.5 (d), 128.8 (d, 2 C), 128.7 (d, 2 C), 127.4 (d), 126.8 (d), 118.2 (d), 113.3 (d), 11.3 (d), 73.6 (d), 55.2 (q), 40.5 (t); HRMS calcd for C₂₃H₂₂O₂S₂ (M⁺) 394.1061, found 394.1070; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 13.7 min (S-isomer), 15.6 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(*m*-(trifluoromethyl)phenyl)-3butenol (11): oil; ¹³C NMR (CDCl₃) δ 144.5 (d), 136.0 (d), 135.7 (s), 133.3 (s), 133.0 (s), 132.1 (d, 2 C), 131.0 (s), 130.3 (d, 2 C), 129.2 (d), 128.9 (d, 3 C), 128.7 (d, 2 C), 127.7 (d), 126.9 (d), 124.3 (d), 122.6 (d), 72.9 (d), 40.5 (t); HRMS calcd for C₂₃H₁₉F₃-OS₂ (M⁺) 432.0829, found 432.0838; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 6.4 min (S-isomer), 10.4 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(p-(trifluoromethyl)phenyl)-3butenol (1m): oil; ¹³C NMR (CDCl₃) δ 147.4 (s), 135.9 (d), 133.7 (s), 133.4 (s), 133.1 (s), 132.2 (d, 2 C), 130.4 (d, 2 C), 126.9 (d), 126.2 (d), 125.5 (d), 125.1 (d), 125.2 (d), 72.9 (d), 40.5 (t); HRMS calcd for C₂₃H₁₉F₃OS₂ (M⁺) 432.0829, found 432.0834; HPLC (Chiracel OD, 2-propanol/hexane (20:80)) $t_{\rm R}$ 12.3 min (S-isomer), 15.8 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(m-fluorophenyl)-3-butenol (1n): oil; 13 C NMR (CDCl₃) δ 164.5 (s), 146.1, 136.6, 136.1, 135.7, 133.9, 133.5, 132.8, 132.1, 130.4, 130.1, 130.0, 128.9, 128.7,

^{(11) (}a) Dziadulewicz, E.; Hodgson, D.; Gallagher, T. J. Chem. Soc., Perkin Trans. 1 1988, 3367. (b) Kolb, M. Synthesis 1990, 171.

128.6, 127.7, 126.9, 121.4, 114.6, 114.3, 112.9, 112.7, 73.0 (d), 40.5 (t); HRMS calcd for $C_{22}H_{19}OS_2F$ (M⁺) 382.0861, found 382.0844; HPLC (Chiracel OD, 2-propanol/hexane (20:80)) t_R 7.4 min (S-isomer), 11.1 min (R-isomer).

4,4-Bis(phenylthio)-1-(p-fluorophenyl)-3-butenol (10): oil; ¹³C NMR (CDCl₃) δ 139.2 (s), 137.2 (d), 133.9 (s), 133.6 (s), 132.4 (s), 131.9 (d, 2 C), 130.4 (d, 2 C), 128.8 (d, 2 C)128.7 (d, 2 C), 127.6 (d), 127.4 (d), 126.8 (d), 115.5 (d), 115.1 (d), 72.9 (d), 40.6 (t); HRMS calcd for C₂₂H₁₉OS₂F (M⁺) 382.0861, found 382.0850; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 9.0 min (S-isomer), 12.5 min (R-isomer).

4.4-Bis(phenylthio)-1-(o-chlorophenyl)-3-butenol (1p): oil; ¹³C NMR (CDCl₃) δ 137.3 (d), 131.9 (d, 4 C), 130.4 (d, 2 C), 129.4 (s, 2 C), 128.8 (d, 2 C), 128.6 (d, 2 C), 127.5 (d, 2 C), 127.3 (s), 127.1 (s), 126.8 (d, 2 C), 69.9 (d), 38.8 (t); HRMS calcd for C₂₂H₁₉OS₂Cl (M⁺) 398.0565, found 398.0567; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 23.6 min (S-isomer), 26.1 min (R-isomer).

4,4-Bis(phenylthio)-1-(m-chlorophenyl)-3-butenol (1q): oil; ¹H NMR (CDCl₃) δ 7.32-7.05 (14 H, m), 6.18 (1 H, t, J = 7.2 Hz), 4.72 (1 H, t, J = 6.4 Hz), 2.86 (2 H, dd, J = 7.2, 6.4 Hz); HRMS calcd for C₂₂H₁₉OS₂Cl (M⁺) 398.0565, found 398.0596; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 12.9 min (S-isomer), 14.9 min (R-isomer).

4,4-Bis(phenylthio)-1-(m-bromophenyl)-3-butenol (1r): oil; ¹³C NMR (CDCl₃, 50 MHz) δ 145.8 (s), 136.2 (d), 135.7 (s), 133.5 (s), 132.9 (s), 132.1 (d, 2 C), 130.7 (d), 130.4 (d), 130.0 (d), 129.0 (d), 128.9 (d, 2 C), 128.8 (d), 128.7 (d, 2 C), 127.7 (d), 126.9 (d), 126.8 (d), 124.5 (d), 122.5 (s), 72.9 (d), 40.4 (t); HRMS calcd for C₂₂H₁₇O⁸¹BrS₂ (M⁺) 441.9900, found 441.9869; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 8.5 min (S-isomer), 13.0 min (R-isomer).

6,6-Bis(phenylthio)-2-methyl-1,5-hexadien-3-ol (1s): oil; ¹³C NMR (CDCl₃, 50 MHz) δ 146.6 (s), 138.7 (d), 135.7 (s), 134.0 (s), 131.7 (d, 2 C), 131.5 (s), 130.4 (d, 2 C), 128.8 (d, 2 C), 128.7 (d, 2 C), 127.4 (d), 126.8 (d), 111.4 (d), 74.8 (d), 36.8 (t), 17.9 (q); HRMS calcd for C₁₉H₂₀OS₂ (M⁺) 328.0956, found 328.0948; HPLC (Chiracel OD, 2-propanol/hexane (20:80)) $t_{\rm R}$ 8.5 min (S-isomer), 13.0 min (*R*-isomer).

5,5-Bis(phenylthio)-4-methyl-4-penten-2-ol (2a): oil; TLC (10% EtOAc in hexane) $R_f = 0.2$; IR (neat) 3383 cm⁻¹; MS m/z (rel intensity) 316 (M⁺, 100), 297 (10), 271 (32), 206 (28), 161 (55), 97 (68); ¹H NMR (CDCl₃) δ 7.25–7.10 (10 H, m), 4.09 (1 H, m), 2.93 (1 H, dd, J = 13.0, 8.3 Hz), 2.68 (1 H, dd, J = 13.0, 5.0 Hz), 2.24 (3 H, s), 1.27 (2 H, d, J = 6.2 Hz); ¹³C NMR (CDCl₃) δ 152.4 (s), 135.4 (s, 2 C), 129.5 (s, 2 C), 128.5 (d, 4 C), 126.2 (s), 126.0 (d), 123.8 (s), 67.3 (d), 46.8 (t), 23.8 (q), 22.9 (q); HRMS calcd for C₁₈H₂₀OS₂ (M⁺) 316.0955, found 316.0963; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 6.5 min (S-isomer), 8.3 min (R-isomer).

6,6-Bis(phenylthio)-5-methyl-5-hexen-3-ol (2b): oil; ¹³C NMR (CDCl₃) δ 153.0 (s), 135.0 (s, 2 C), 129.4 (d, 2 C), 129.2 (d, 2 C), 128.6 (d, 4 C), 126.3 (d), 126.1 (d), 123.3 (s), 72.5 (d), 44.9 (t), 30.9 (t), 23.0 (q), 10.0 (q); HRMS calcd for C₁₉H₂₂OS₂ (M⁺) 330.1112, found 330.1180; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 9.1 min (S-isomer), 11.3 min (R-isomer).

1,1-Bis(phenylthio)-2-methyl-1-hepten-4-ol (2c): oil; ¹³C NMR (CDCl₃) δ 153.0 (s), 135.5 (s, 2 C), 129.4 (d, 2 C), 129.1 (d, 2 C), 128.6 (d, 4 C), 126.2 (d), 126.0 (d), 123.7 (s), 70.9 (d), 45.3 (t), 40.2 (t), 23.0 (q), 18.8 (q), 14.0 (q); HRMS calcd for C₂₀H₂₄OS₂ (M⁺) 344.1268, found 344.1263; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 6.4 min (S-isomer), 9.3 min (*R*-isomer).

6,6-Bis(phenylthio)-2,5-dimethyl-5-hexen-3-ol (2d). Oil; ¹³C NMR (CDCl₃) δ 154.0 (s), 135.5 (s, 2 C), 130.1 (d, 2 C), 129.7 (d, 2 C), 129.3 (d, 4 C), 126.9 (d), 126.7 (d), 123.1 (s), 78.2 (d), 42.9 (t), 35.0 (d), 23.5 (q), 19.4 (q), 17.9 (q); HRMS calcd for C₂₀H₂₄OS₂ (M⁺) 344.1265, found 344.1263; HPLC (Chiracel OD, 2-propanol/hexane (3:97)) $t_{\rm R}$ 13.4 min (*R*-isomer), 17.0 min (*S*-isomer).

4,4-Bis(phenylthio)-3-methyl-1-phenyl-3-butenol (2e): oil; ¹³C NMR (CDCl₃) δ 151.8 (s), 143.8 (s), 135.5 (s), 135.4 (s), 129.7 (d, 2 C), 129.1 (d, 2 C), 128.6(d, 6 C), 127.0 (d, 2 C), 126.3-(d), 126.0 (d), 125.9 (d), 123.8 (s), 73.7 (d), 46.8 (t), 23.0 (q); HRMS calcd for C₂₃H₂₂OS₂ (M⁺) 378.1112, found 378.1086; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 9.3 min (*R*-isomer), 14.0 min (*S*-isomer).

1,1-Bis(phenylthio)-2-methyl-1-octen-4-ol (2f): oil; ¹³C NMR (CDCl₃) δ 153.1 (s), 135.6 (s), 135.5 (s), 129.5 (d, 2 C), 129.1 (d, 2 C), 128.6 (d, 4 C), 126.2 (d), 126.0 (d), 123.7 (s), 71.1 (d), 45.4 (t), 37.7 (t), 27.8 (t), 23.0 (q), 22.6 (t), 14.0 (q); HRMS calcd for C₂₁H₂₆OS₂ (M⁺) 358.1425, found 358.1419; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 5.7 min (*S*-isomer), 7.5 min (*R*-isomer).

1,1-Bis(phenylthio)-2-methyl-1-nonen-4-ol (2g). Oil; ¹³C NMR (CDCl₃) δ 153.0 (s), 135.6 (s, 2 C), 129.4 (d, 2 C), 128.6 (d, 4 C), 126.2 (d), 126.0 (d), 123.6 (s), 71.1 (d), 45.4 (t), 38.0 (t), 31.7 (t), 25.3 (t), 22.9 (t), 22.5 (q), 14.0 (q); HRMS calcd for C₂₂H₂₈OS₂ (M⁺) 372,1581, found 372.1586; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 4.7 min (S-isomer), 6.4 min (*R*-isomer).

1-(1,3-Dithianylidene)-2-methyl-3-pentanol (3a): HPLC (Chiracel OD, 2-propanol/hexane (1:99)) $t_{\rm R}$ 18.6 min (2S,3*R*-isomer), 20.8 min (2*R*,3*S*-isomer).

1-(1,3-Dithianylidene)-2-methyl-3-heptanol (3b): HPLC (Chiracel OD, 2-propanol/hexane (1:99)) $t_{\rm R}$ 13.1 min (2S,3Risomer), 14.7 min (2R,3S-isomer).

1-(1,3-Dithianylidene)-2-methyl-3-octanol (3c): HPLC (Chiracel OD, 2-propanol/hexane (1:99)) $t_{\rm R}$ 12.8 min (2S,3*R*-isomer), 14.6 min (2*R*,3*S*-isomer).

1-(1,3-Dithianylidene)-2-methyl-3-nonanol (3d): HPLC (Chiracel OD, 2-propanol/hexane (1:99)) $t_{\rm R}$ 11.4 min (2S,3*R*-isomer), 13.3 min (2*R*,3*S*-isomer).

5,5-Bis(phenylthio)-4-penten-2-yl acetate (7a): oil; TLC (10% EtOAc in hexane) $R_f = 0.4$; IR (neat) 1731 cm⁻¹; MS m/z (rel intensity) 345 (M⁺+1, 12), 344 (1), 284 (100), 207 (50); ¹H NMR (CDCl₃) δ 7.33–7.16 (10 H, m), 6.23 (1 H, t, J = 7.4 Hz), 4.99 (1 H, m), 2.71 (2 H, dd, J = 6.4, 6.4 Hz), 2.01 (3 H, s), 1.24 (3 H, d, J = 6.3 Hz); ¹³C NMR (CDCl₃) δ 170.4 (s), 137.0, 133.9, 133.7, 132.5, 132.0, 130.5 (d, 2 C), 128.9 (d, 2 C), 128.7 (d, 2 C), 127.5 (d), 126.9 (d), 125.8, 69.8 (d), 37.3 (t), 21.2 (q), 19.7 (q); HRMS calcd for C₁₉H₂₀O₂S₂ (M⁺) 344.0904, found 344.0909.

6,6-Bis(phenylthio)-5-hexen-3-yl acetate (7b): oil; IR (neat) 1724 cm⁻¹; 13 C NMR (CDCl₃) δ 170.6 (s), 137.2 (d), 133.9 (s), 133.7 (s), 132.2 (s), 131.9 (d, 2 C), 130.5 (d, 2 C), 128.8 (d, 2 C), 128.6 (d, 2 C), 127.6 (d), 126.8 (d), 74.3 (d), 35.2 (t), 26.8 (t), 21.2 (q), 9.6 (q); HRMS calcd for C₂₀H₂₂O₂S₂ (M⁺) 358.1061, found 358.1034.

1,1-Bis(phenylthio)-1-hepten-4-yl acetate (7c): oil; IR (neat) 1730 cm⁻¹; ¹³C NMR (CDCl₃, 50 MHz) δ 170.6 (s), 137.3 (d), 133.9 (s), 133.1 (s), 132.3 (s), 132.0 (d, 2 C), 130.5 (d, 2 C), 128.8 (d, 2 C), 128.7 (d, 2 C), 127.6 (d), 126.9 (d), 72.9 (d), 36.0 (t), 35.7 d), 21.2 (q), 18.5 (q), 13.9 (q); HRMS calcd for C₂₁H₂₄O₂S₂ (M⁺) 372.1217, found 372.1225

6,6-Bis(phenylthio)-2-methyl-5-hexen-3-yl acetate (7d): oil; ¹³C NMR (CDCl₃) δ 170.6 (s), 137.7 (d), 134.0 (s, 2 C), 133.8 (s), 132.1 (d, 2 C), 130.5 (d, 2 C), 128.9 (d, 2 C), 128.7 (d, 2 C), 127.6 (d), 126.9 (d), 76.5 (d), 33.2 (t), 31.5 (d), 21.1 (q), 18.7 (q), 17.8 (q); HRMS calcd for C₂₁H₂₄O₂S₂ (M⁺) 372.1217, found 372.1198

4,4-Bis(phenylthio)-1-phenyl-3-butenyl acetate (7e): oil; IR (neat) 1724 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.8 (s), 139.4 (s), 135.9 (d), 133.7 (s), 133.4 (s), 132.9 (s), 132.0 (d, 2 C), 130.4 (d, 2 C), 128.7 (d, 2 C), 128.6 (d, 2 C), 128.4 (d, 2 C), 128.0 (d, 2 C), 127.5 (d), 126.8 (d), 126.3 (d, 2 C), 74.5 (d), 37.7 (t), 21.0 (q); HRMS calcd for C₂₄H₂₂O₂S₂ (M⁺) 406.1061, found 406.1069.

4,4-Bis(phenylthio)-1-(o-methylphenyl)-3-butenyl acetate (7f): oil; IR (neat) 1727 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.9 (s), 138.0 (s), 135.9 (d), 133.8 (s), 133.8 (s), 133.4 (s), 133.0 (s), 132.2 (d, 2 C), 130.4 (d, 2 C), 128.8 (d, 2 C), 128.7 (d, 2 C), 127.8 (d), 127.7 (d), 126.8 (d), 126.1 (d), 125.8 (d), 71.6 (d), 37.0 (t), 21.1 (q), 19.1 (q); HRMS calcd for C₂₅H₂₄O₂S₂ (M⁺) 420.1217, found 420.1216.

4,4-Bis(phenylthio)-1-(*m*-methylphenyl)-3-butenyl acetate (7g): oil; IR (neat) 1733 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.7 (s), 139.3 (s), 137.9 (s), 136.2 (d), 133.7 (s), 133.4 (s), 132.6 (s), 131.8 (d, 2 C), 130.3 (d, 2 C), 128.7 (d, 3 C), 128.5 (d, 2 C), 128.2 (d), 127.4 (d), 126.9 (d), 123.3 (d), 74.5 (d), 37.7 (t), 21.3 (q), 21.0 (q); HRMS calcd for C₂₅H₂₄O₂S₂ (M⁺) 420.1217, found 420.1216.

4,4-Bis(phenylthio)-1-(*p*-methylphenyl)-3-butenyl acetate (7h): oil; IR(neat) 1729 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.0 (s), 137.7 (s), 136.5 (s), 136.4 (d), 133.8 (s), 133.5 (s), 132.7 (s), 131.9 (d, 2 C), 130.5 (d, 2 C), 129.1 (d, 2 C), 128.8 (d, 2 C), 128.6 (d, 2 C), 127.5 (d), 126.8 (d), 126.4 (d, 2 C), 74.5 (d), 37.7 (t), 21.1 (q); HRMS calcd for C₂₅H₂₄O₂S₂ (M⁺) 420.1217, found 420.1216.

4,4-Bis(phenylthio)-1-(*p*-(methylethyl)phenyl)-3-butenyl acetate (7i): oil; IR (neat) 1734 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.0 (s), 148.7 (s). 136.9 (s), 136.2 (d), 133.8 (s), 133.5 (s), 132.8 (s), 132.0 (d, 2 C), 130.5 (d, 2 C), 128.8 (d, 2 C), 127.6 (d), 126.9 (d), 126.5 (d, 4 C), 74.6 (d), 37.8 (t), 33.7 (d), 23.9 (q, 2 C), 21.2 (q); HRMS calcd for C₂₇H₂₈O₂S₂ (M⁺) 448.1530, found 448.1523; HPLC (Chiracel OD, 2-propanol/hexane (20:80)) $t_{\rm R}$ 4.7 min (S-isomer), 5.2 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(o-methoxyphenyl)-3-butenyl acetate (7j): oil; IR (neat) 1737 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.6 (s), 155.8 (s), 137.6(d), 134.0 (s), 133.7 (s), 131.9 (s), 131.6 (d, 2 C), 130.2 (d, 2 C), 128.6 (d, 3 C), 128.5 (d, 2 C), 127.9 (d), 127.2 (d), 126.6 (d), 126.1 (d), 120.3 (d), 120.3 (d), 110.4 (d), 69.3 (d), 55.2 (q), 36.5 (t), 21.0 (q); HRMS calcd for C₂₅H₂₄O₃S₂ (M⁺) 436.1167, found 436.1165; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 5.8 min (S-isomer), 7.9 min (R-isomer).

4,4-Bis(phenylthio)-1-(*m*-methoxyphenyl)-3-butenyl acetate (7k): oil; IR (neat) 1732 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.9 (s), 156.0 (s), 137.8 (d), 134.1 (s), 133.9 (s), 132.0 (s), 131.8 (d, 2 C), 130.4 (d, 2 C), 128.7 (d, 2 C), 128.6 (d, 2 C), 128.1 (s), 127.4 (d), 127.3 (d), 126.7 (d), 126.3 (d), 120.5 (d), 110.5 (d), 69.5(d), 55.4 (q), 36.6 (t), 21.2 (q); HRMS calcd for C₂₅H₂₄O₃S₂ (M⁺) 436.1167, found 436.1160.

4,4-Bis(phenylthio)-1-(*m*-(trifluoromethyl)phenyl)-3butenyl acetate (71): oil; IR (neat) 1730 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.8 (s), 141.1 (d), 134.4, 133.8, 132.3, 130.4, 130.0, 129.8, 129.2, 129.1, 128.9, 128.7, 128.5, 128.2, 127.8, 126.9, 126.5, 124.6, 73.9 (d), 37.6 (t), 21.0 (q); HRMS calcd for C₂₄H₂₁F₃O₂S₂ (M⁺) 474.0935, found 474.0934; HPLC (Chiracel OD, 2-propanol/hexane (10:90)) $t_{\rm R}$ 3.6 min (S-isomer), 3.8 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(*p*-(trifluoromethyl)phenyl)-3butenyl acetate (7m): oil; IR (neat) 1733 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.9 (s), 143.6 (s), 134.1 (d), 134.0 (s), 133.5 (s), 133.1 (s), 132.4 (d, 2 C), 130.5 (d, 2 C), 129.9 (s), 128.9 (d, 2 C), 128.7 (d, 2 C), 127.9 (d), 127.0 (d), 126.7 (d), 125.5 (d, 2 C), 125.4 (d), 74.0 (d), 37.6 (t), 21.0 (q); HRMS calcd for C₂₅H₂₁O₂S₂F₃ (M⁺) 474.0935, found 474.0934.

4,4-Bis(phenylthio)-1-(m-fluorophenyl)-3-butenyl acetate (7n): oil; IR (neat) 1731 cm⁻¹; MS m/z (rel intensity) 424 (M⁺, 8), 365 (50), 257 (100); ¹H NMR (CDCl₃) δ 7.30–7.05 (14 H, m), 6.04 (1 H, t, J = 7.4 Hz), 5.79 (1 H, t, J = 6.3 Hz), 2.99 (2 H, dd, J = 7.4, 6.3 Hz), 2.09 (3 H, s); HRMS calcd for C₂₄H₂₁O₂S₂F (M⁺) 424.0967, found 424.0972; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 9.1 min (S-isomer), 10.7 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(p-fluorophenyl)-3-butenyl acetate (70): oil; IR (neat) 1731 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.9 (s), 164.0, 160.8, 135.4 (s), 135.3 (d), 133.7 (s), 133.4 (s), 132.2 (d, 2 C), 130.5 (d), 130.4 (d, 2 C), 128.8 (d, 2 C), 128.7 (d, 2 C), 128.3 (d), 128.2 (d), 127.5 (d), 126.9 (d), 115.5 (d), 115.2 (d), 74.0 (d), 37.7 (t), 21.1 (q); HRMS calcd for C₁₈H₂₀OS₂ (M⁺) 424.0967, found 424.0973; HPLC (Chiracel OD, 2-propanol/hexane (3:97)) $t_{\rm R}$ 8.8 min (S-isomer), 10.2 min (*R*-isomer).

4,4-Bis(phenylthio)-1-(o-chlorophenyl)-3-butenyl acetate (7p): oil; IR (neat) 1738 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.5 (s), 137.4 (s), 135.2 (d), 133.7 (s), 133.4 (s), 133.3 (d), 132.2 (d), 132.0 (s), 130.4 (d, 2 C), 129.5 (d), 128.9 (d), 128.8 (d, 2 C), 128.6 (d, 2 C), 127.6 (d), 127.1 (d), 126.9 (d), 126.8 (d), 71.3 (d), 36.4 (t), 21.0 (q); HRMS calcd for C₂₄H₂₁O₂S₂Cl (M⁺) 440.0671, found 440.0663; HPLC (Chiracel OD, 2-propanol/ hexane (10:90)) $t_{\rm R}$ 3.6 min (S-isomer), 3.8 min (R-isomer).

4,4-Bis(phenylthio)-1-(*m*-chlorophenyl)-3-butenyl acetate (7q): oil; IR (neat) 1736 cm⁻¹; 13 C NMR (CDCl₃, 50 MHz) δ 169.7 (s), 141.6 (s), 134.4 (d), 134.3 (s), 133.7 (s), 133.6 (s), 133.1 (s), 132.3 (d, 2 C), 130.4 (d, 2 C), 129.7 (d), 128.8 (d, 2 C), 128.6 (d, 2 C), 128.1 (d), 127.7 (d), 126.9 (d), 126.4 (d), 124.5 (d), 73.8 (d), 37.5 (d), 20.9 (q); HRMS calcd for C₂₄H₂₁O₂S₂Cl

4,4-Bis(phenylthio)-1-(*m*-bromophenyl)-3-butenyl acetate (7r): oil; ¹³C NMR (CDCl₃) δ 169.7 (s), 141.9 (s), 134.3 (d), 133.7 (s), 133.6 (s), 133.1 (s), 132.3 (d), 131.1 (d, 2 C), 130.4 (d, 2 C), 130.0 (d), 129.3 (d), 128.9 (d, 2 C), 128.7 (d, 2 C), 127.8 (d), 126.9 (d), 125.0 (d), 122.5 (s), 73.8 (d), 37.6 (t), 21.0 (q); HRMS calcd for C₂₄H₂₁O₂⁷⁹BrS₂ (M⁺) 484.0166, found 484.0151; HPLC (Chiracel OD, 2-propanol/hexane (5:95)) $t_{\rm R}$ 8.1 min (S-isomer), 8.9 min (*R*-isomer).

6,6-Bis(phenylthio)-2-methyl-1,5-hexadien-3-yl acetate (7s): oil; IR (neat) 1734 cm⁻¹; ¹³C NMR (CDCl₃) δ 169.9 (s), 142.3 (s), 136.6 (d), 133.9 (s), 133.6 (s), 132.4 (s), 132.0 (d, 2 C), 128.8 (d, 2 C), 128.6 (d, 2 C), 127.5 (d), 126.8 (d), 112.9 (t), 75.7 (d), 34.3 (t), 21.0 (q), 18.4 (q); HRMS calcd for $C_{21}H_{22}O_2S_2$ (M⁺) 370.1061, found 370.1025.

5,5-Bis(phenylthio)-4-methyl-4-penten-2-yl acetate (8a): oil; TLC (5% EtOAc in hexane) $R_f = 0.30$; IR (neat) 1730 cm⁻¹; MS m/z (rel intensity) 358 (M⁺, 25), 298 (65), 221 (100); ¹H NMR (CDCl₃) δ 7.24–7.05 (10 H, m), 5.23 (1 H, m), 3.08 (1 H, dd, J = 13.2, 8.7 Hz), 2.74 (1 H, dd, J = 13.2, 4.7 Hz), 2.20 (3 H, s), 2.00 (3 H, s), 1.29 (3 H, d, J = 6.2 Hz); ¹³C NMR (CDCl₃) δ 170.3 (s), 150.3 (s), 135.2 (s, 2 C), 129.7 (d, 2 C), 129.6 (d, 2 C), 128.4 (d, 4 C), 126.3 (d), 126.2 (d), 125.2 (s), 69.3 (d), 44.0 (t), 22.5 (q), 21.2 (q), 20.2 (q); HRMS calcd for C₂₀H₂₂O₂S₂ (M⁺) 358.1061, found 358.1041.

6,6-Bis(phenylthio)-5-methyl-5-hexen-3-yl acetate (8b): oil; IR (neat) 1731 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.4 (s), 150.4 (s), 135.3 (s, 2 C), 129.8 (d, 4 C), 128.5 (d, 4 C), 126.3 (d, 2 C), 125.2 (s), 73.7 (d), 41.7 (t), 27.5 (t), 22.6 (q), 21.1 (q), 9.6 (q); HRMS calcd for C₂₁H₂₄O₂S₂ (M⁺) 372.1217, found 372.1204.

1,1-Bis(phenylthio)-2-methyl-1-hepten-4-yl acetate (8c): oil; IR (neat) 1732 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.5 (s), 150.6 (s), 135.4 (s, 2 C), 129.9 (d, 2 C), 129.8 (d, 2 C), 128.5 (d, 4 C), 126.3 (d, 2 C), 125.2 (s), 72.4 (d), 42.2 (d), 36.8 (t), 22.7 (q), 21.2 (q), 18.6 (t), 13.8 (q); HRMS calcd for C₂₂H₂₆O₂S₂ (M⁺) 386.1374, found 386.1374.

6,6-Bis(phenylthio)-2,5-dimethyl-5-hexen-3-yl acetate (8d): oil; IR (neat) 1726 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.3 (s), 150.6 (s), 135.3 (s), 135.2 (s), 129.9 (d, 2 C), 129.7 (d, 2 C), 128.4 (d, 4 C), 126.2 (d), 126.17 (d), 125.0 (s), 76.5 (d), 39.2 (t), 32.2 (d), 22.6 (q), 21.0 (q), 18.3 (q), 17.6 (q); HRMS calcd for C₂₃H₂₄OS₂ (M⁺) 386.1374, found 386.1376.

4,4-Bis(phenylthio)-3-methyl-1-phenyl-3-butenyl acetate (8e): oil; IR (neat) 1736 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.0 (s), 149.5 (s), 139.7 (s), 135.3 (s), 135.2 (s), 130.0 (d, 2 C), 129.5 (d, 2 C), 128.5 (d, 4 C), 128.1 (d), 126.6 (d, 2 C), 126.4 (d), 126.2 (d), 126.0 (s), 74.5 (d), 44.0 (t), 22.6 (q), 21.2 (q); HRMS calcd for C₂₅H₂₄O₂S₂ (M⁺) 420.1218, found 420.1224.

1,1-Bis(phenylthio)-2-methyl-1-octen-4-yl acetate (8f): oil; IR (neat) 1733 cm⁻¹; 13 C NMR (CDCl₃) δ 170.3 (s), 150.4 (s), 135.3 (s), 135.2 (s), 129.7 (d, 2 C), 129.68 (d, 2 C), 128.4 (d, 4 C), 126.2 (d, 2 C), 125.2 (s), 72.5 (d), 42.0 (t), 34.3 (t), 27.3 (t), 22.5 (q), 22.3 (t), 21.1 (q), 13.8 (q); HRMS calcd for C₂₃H₂₈O₂S₂ (M⁺) 440.1530, found 400.1536.

1,1-Bis(phenylthio)-2-methyl-1-nonen-4-yl acetate (8g): oil; IR (neat) 1731 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.6 (s),150.7 (s), 135.5 (s), 134.4 (s), 129.9 (d, 2 C), 129.8 (d, 2 C), 128.6 (d, 4 C), 126.4 (d, 2 C), 125.6 (s), 72.8 (d), 42.2 (t), 34.7 (t), 31.6 (t), 25.0 (t), 22.7 (q), 22.5 (t), 21.3 (q), 14.0 (q); HRMS calcd for C₂₄H₃₀O₂S₂ (M⁺) 414.1687, found 414.1677.

1-(1,3-Dithianylidene)-2-methyl-3-pentyl acetate (9a): oil; TLC (5% EtOAc in hexane) $R_f = 0.3$; IR (neat) 1731 cm⁻¹; MS m/z (rel intensity) 261 (M⁺+1, 18), 260 (1), 200 (30), 159 (100); ¹H NMR (CDCl₃) δ 5.75 (1 H, d, J = 9.8 Hz), 4.69 (1 H, m), 2.87 (1H, m), 2.84 - 2.74 (4 H, m), 2.13 - 2.04 (2 H, m), 1.98 (3 H, s), 1.47 (2 H, m), 0.91 (3 H, d₇J = 4.2 Hz), 0.78 (3 H, t, J = 5.0 Hz); ¹³C NMR (CDCl₃) δ 170.7 (s), 134.5 (d), 127.3 (s), 78.1 (s), 37.3 (t), 30.2 (t), 29.6 (t), 25.2 (t), 25.1 (t), 20.9 (q), 16.7 (q), 9.8 (q); HRMS calcd for C₁₃H₂₂O₂S₂ (M⁺) 260.0904, found 260.0912.

1-(1,3-Dithianylidene)-2-methyl-3-heptyl acetate (9b): oil; IR (neat) 1730 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.8 (s), 134.7 (d), 127.3 (s), 76.8 (d), 37.3 (d), 31.9 (t), 30.3 (t), 29.7 (t), 27.6 (t), 25.1 (t), 22.5 (t), 21.0 (q), 16.7 (q), 13.9 (q); HRMS calcd for C₁₄H₂₄O₂S₂ (M⁺) 288.1217, found 288.1216. **1-(1,3-Dithianylidene)-2-methyl-3-octyl acetate (9c):** oil; IR (neat) 1731 cm⁻¹; ¹³C NMR (CDCl₃) δ 170.6 (s), 134.5 (d), 127.2 (s), 76.7 (d), 37.6 (d), 32.1 (t), 31.5 (t), 30.2 (t), 29.6 (t), 25.0 (t), 24.9 (t), 22.3 (t), 20.9 (q), 16.7 (q), 13.8 (q); HRMS calcd for $C_{15}H_{26}O_2S_2$ (M⁺) 302.1374, found 302.1348.

1-(1,3-Dithianylidene)-2-methyl-3-nonyl acetate (9d): oil; IR (neat) 1731 cm⁻¹; ¹³C NMR (CDCl₃, 50 MHz) δ 170.6 (s), 134.6 (d), 127.2 (s), 76.9 (d), 37.6 (d), 32.2 (t), 31.6 (t), 30.2 (t), 29.6 (t), 29.0 (t), 25.3 (t), 25.0 (t), 22.4 (t), 20.9 (q), 16.7 (q), 14.0 (q); HRMS calcd for $C_{16}H_{28}O_2S_2$ (M⁺) 316.1530, found 316.1530.

(S)-4-Hydroxy-2-heptanone (10). A cold (-78 °C) solution of (S)-2c (0.18 g, 0.47 mmol, 86% ee) in CH₂Cl₂ (30 mL) was treated with ozone. The reaction was monitored by TLC analysis. After complete consumption of (S)-8c, Me₂S (0.5 mL) was added. The mixture was warmed to room temperature and stirring maintained for 4 h. The mixture was concentrated and separated by silica gel chromatography and HPLC (EtOAc/hexane (1:1)) to give (S)-10 (20 mg, 38%), $[\alpha]^{25}_{\rm D}$ +44° (c = 0.18, CHCl₃) (lit.⁷ $[\alpha]_{\rm D}$ +35.1° (c = 2.1, CHCl₃, 58% ee)). Two isomers of 3-hydroxy-3-methyl-5-propyl-2,3,4,5-tetrahydrofuran-2-one (37%, 55:45) were also isolated as side products.

5-Phenyl-2,3,4,5-tetrahydrofuran-2-one (11e). The alcohol **1e** (60 mg, 91% ee favoring (*R*)-configuration), obtained from lipase-catalyzed hydrolysis of acetate **7e** (entry 6 of Table 2), was treated with HgCl₂ by a procedure silimar to that for hop lactone to give (+)-**11e** (24 mg, 90%) in favor of the (*R*)configuration (91% ee), $[\alpha]^{25}_{D} + 18^{\circ}$ (c = 1.1, CHCl₃) (lit.⁸ $[\alpha]_{D}$ +32.5° (c = 1, MeOH)).

5-(Methylethenyl)-2,3,4,5-tetrahydrofuran-2-one (Hop Lactone) (11s). (S)-Alcohol 1s (0.44 g, 1.34 mmol) with 95% ee, obtained by saponification of the recovered acetate (S)-7s (95% ee) in entry 24 of Table 2, was treated with HgCl₂ (0.39 g, 1.47 mmol) in 10% MeOH (10 mL) at reflux for 5 h. The mixture was concentrated and extracted three times with EtOAc. The combined extracts were washed with brine, dried (Na₂SO₄), concentrated, and separated by silica gel chromatography (EtOAc/hexane (1:9)) to give hop lactone (0.15 g, 85%) with $[\alpha]^{25}_{\rm D}$ +10° (c = 1.5, EtOH). Hop lactone in nature has been reported to have optical rotation $[\alpha]_{\rm D}$ +8° (c = 0.25, EtOH).¹ (*R*)-1s (75% ee, entry 24 of Table 2)) was similarly converted to (*R*)-hop lactone, $[\alpha]^{25}_{\rm D}$ -6.5° (c = 2.6, EtOH).

5-Butyl-4-methyl-2,3,4,5-tetrahydrofuran-2-one (Quercus Lactone) (12b). The alcohol 3b (0.22 g, 85% ee favoring the (2S,3R)-configuration), obtained from lipase-catalyzed hydrolysis of acetate 9b, was treated with HgCl₂ by a procedure similar to that for hop lactone to give trans quercus lactone (153 mg, 92%) in favor of (4S,5R)-configuration (85% ee), $[\alpha]^{25}_{\rm D}$ +68° (c = 1.2, MeOH) (lit.² $[\alpha]_{\rm D}$ +79° (c = 1.04, MeOH)). Hydrolysis of (2R,3S)-3b (95% ee) gave (4R,5S)-quercus lactone (95% ee), $[\alpha]^{25}_{\rm D}$ -75° (c = 1.0, MeOH).

5-Pentyl-4-methyl-2,3,4,5-tetrahydrofuran-2-one (Cognac Lactone) (12c). The alcohol 3c (100 mg, 94% ee favoring the (2S,3R)-configuration), obtained from lipasecatalyzed hydrolysis of acetate 9c, was treated with HgCl₂ by a procedure silimar to that for hop lactone to give trans cognac lactone (65 mg, 92%) in favor of (4S,5R)-configuration (96% ee), $[\alpha]^{25}_{D} + 75^{\circ}$ (c = 1.0, CH₂Cl₂), lit.³ $[\alpha]_{D} + 79.5^{\circ}$ (c = 1.12, CH₂Cl₂). Hydrolysis of (2R,3S)-3c (37% ee) gave (4R,5S)cognac lactone, $[\alpha]_{D}^{25} - 35^{\circ}$ (c = 2.9, CH₂Cl₂).

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Supplementary Material Available: NMR spectra and additional spectral data of new compounds (65 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; ordering information is given on any current masthead page.